The Influence of Substrate Surface Chemistry on Biofilm Formation

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Introduction

Protein-mediated adsorption of polysaccharides to surfaces is a critical step in the early stages of biofilm formation. This work focuses on the role of the substrate in this process of biofilm nucleation.

Alginate is a linear, charged polysaccharide composed of mannuronic and glucuronic acid subunits. Its relative structural simplicity makes it ideal for exploring the electrostatic interactions that play an important role in polysaccharide/surface interactions. Positively-charged (lysine-rich) proteins such as Mussel Adhesive Protein are known to promote adsorption of alginate, as is the polypeptide poly-L-lysine.¹

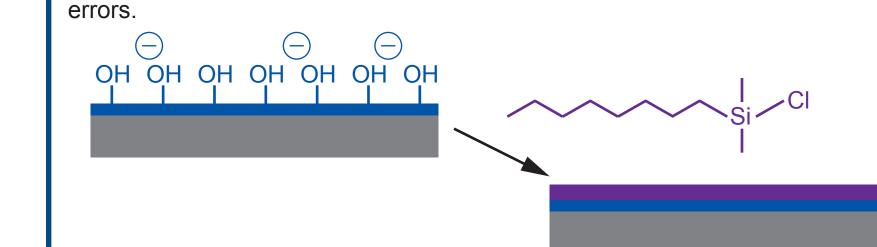
In this work we focus on the role of the surface chemistry of the underlying substrate as it influences poly-L-lysine (PLL) mediated adsorption of alginate. We focus on two representative surfaces: a hydrophobic, electrically neutral surface and a hydrophilic, negatively charged one. The effect of these different surface properties on initial PLL adsorption and on the subsequent adsorption of alginate is investigated with a combination of ellipsometry, dynamic contant angle goniometry and atomic force

Experimental

Hydrophilic silica surfaces were prepared by piranha clean of native oxideterminated Si(100). Hydrophobic silane layers were prepared by vapor phase silanization of these substrates with octyldimethylchlorosilane (Gelest). PLL (Sigma Aldrich) was adsorbed to the hydrophobic surfaces from pH 11 buffer² and to the hydrophilic surfaces from aqueous KBr solutions of varying ionic strength. Alginate (Sigma Aldrich) was adsorbed from aqueous solution. Samples were rinsed with DI water and dried with compressed air prior to measurement.

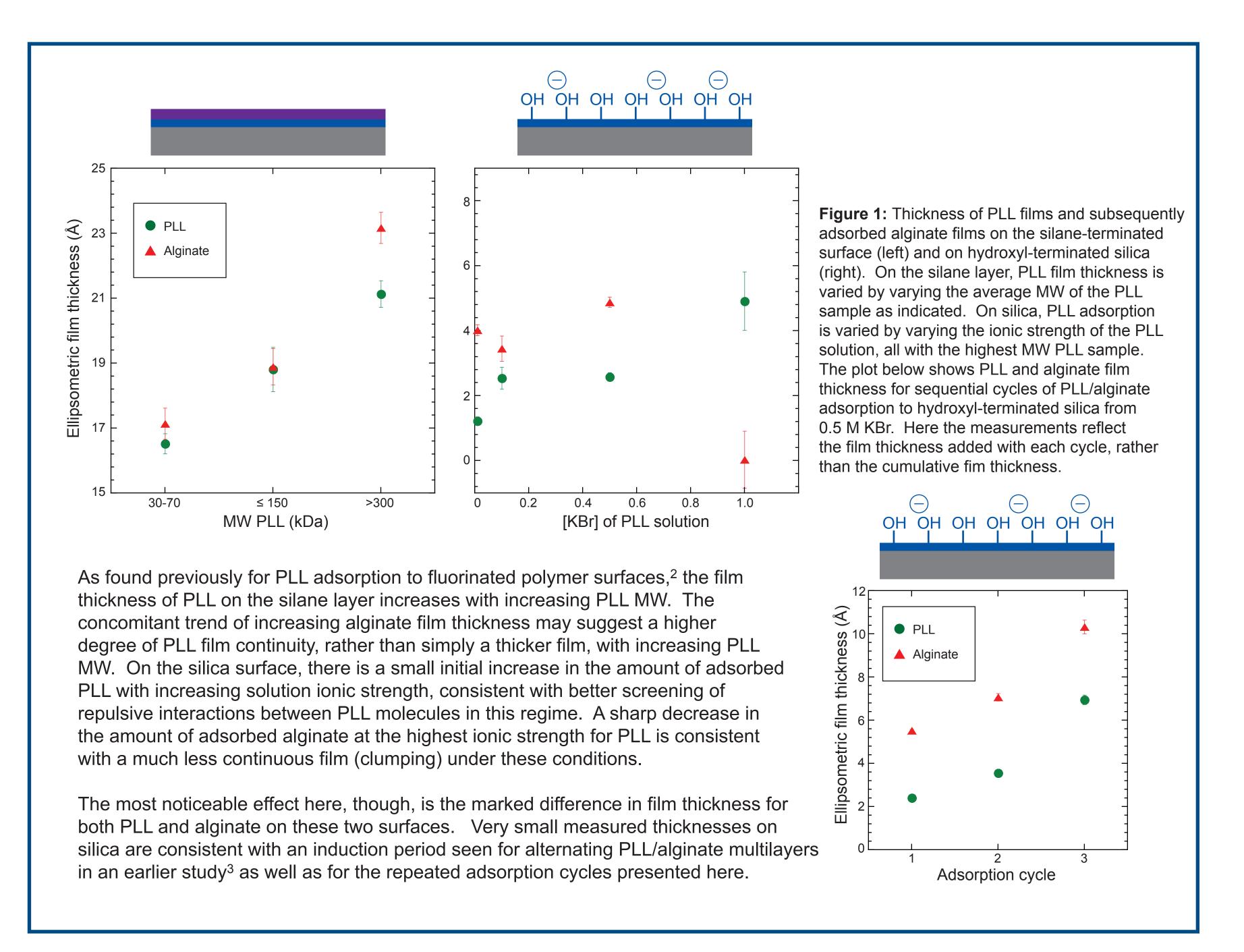
Film thickness was measured with a Gaertner Scientific L116S300 Stokes Ellipsometer at 70°. Advancing and receding contact angles were recorded with a Ramé-Hart manual goniometer. Each sample was measured 3-5 times, and each experiment reported was repeated 4-20 times.

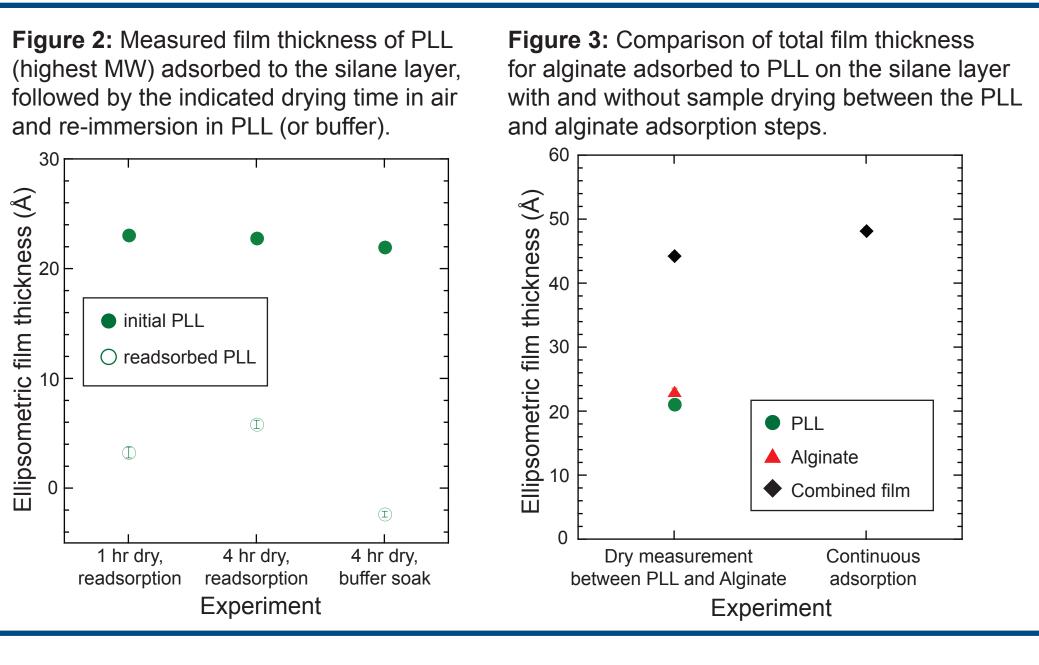
Since multiple measurements were made on each sample, the errors around each estimate were not independent, and the usual way of estimating and determining standard erros is invalid. Instead, estimates and their standard errors (represented by error bars in all graphs) came from a fit of a repeated measures model that includes error terms for both within and across sample



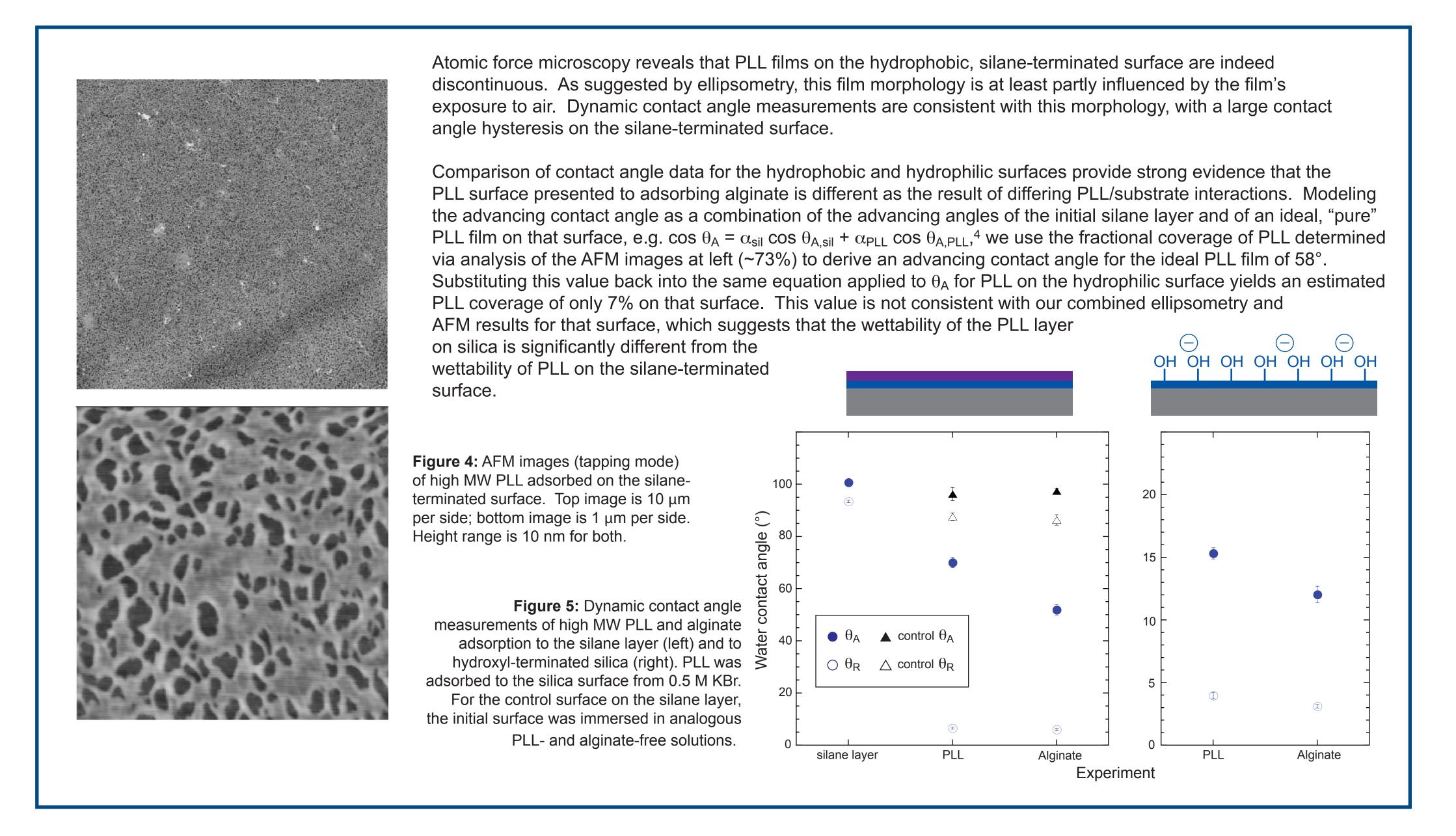
References

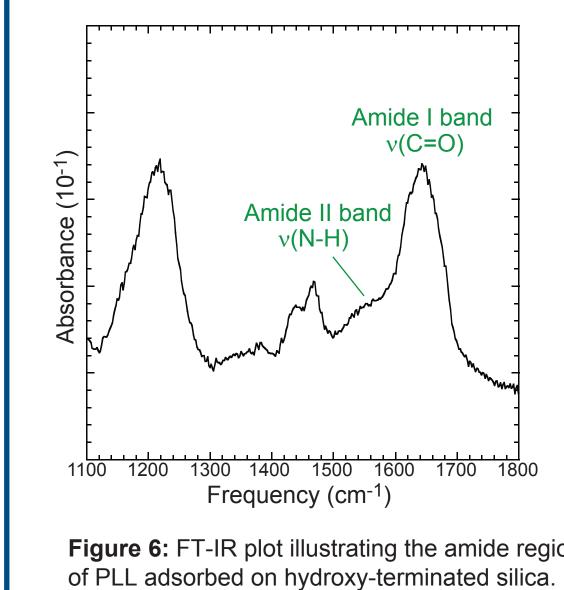
1) P. A. Suci and G.G. Geesey, *J. Colloid Interface Sci.* **1995**, *172*, 347-357. 2) M. S. Shoichet and T. J. McCarthy, *Macromolecules* **1991**, *24*, 1441-1442. 3) D. L. Elbet, C. B. Herbert and J. A. Hubbell, *Langmuir* **1999**, *15*, 5355-5362. 4) C. W. Extrand, *Langmuir* **2003**, *19*, 3793-3796 and references therein.





Measuring PLL and alginate film thickness separately requires drying the sample between adsorption steps. As Figure 2 shows, allowing the PLL film on the silane layer to sit in air causes changes in the film that permit further PLL adsorption upon reimmersion in PLL solution. The data in Figure 3 show a small but significant increase in total (PLL plus alginate) film thickness when this drying step is eliminated. Since alginate adsorption is correlated with the amount of PLL adsorbed on this substrate, this is consistent with PLL films that are slightly more continuous when hydrated.





Alginate adsorption to PLL films on the hydrophobic surface causes a marked change in the observed film morphology. An increase in film height is consistent with ellipsometry results that suggest alginate adsorbs only to PLL-covered regions of the surface. Restructuring of the film indicates a strong interaction between alginate and the underlying PLL. This type of restructuring is not seen for alginate adsorption to PLL on hydroxyl-terminated silica, where the PLL and substrate are expected to interact more strongly through electrostatic attraction. Studies are underway to Figure 6: FT-IR plot illustrating the amide region monitor changes in the conformation of adsorbed PLL via in-situ infrared spectroscopy of these systems.

Conclusions

Interactions between poly-L-lysine and the underlying substrate govern not only the adsorption of PLL itself but also the subsequent adsorption of alginate to the PLL-coated surfaces in this study. While protein-mediated adsorption of alginate is essential (alginate does not adsorb to either of these surfaces in the absence of pre-adsorbed PLL or another protein conditioning film), this mediation is highly dependent on interactions between PLL and the surface. Strong interactions between PLL and negativelycharged silica result in flat PLL films on that surface. On a silane-terminated surface PLL adsorbs in much thicker films whose morphology appears to be dictated by PLL/PLL interactions and, upon subsequent adsorption, to PLL/alginate interactions. Quantitative comparison of the wettabilities of films on these two surfaces provides information about the PLL surface presented to incoming adsorbates, an effective method for investigating the relative properties of films composed of large, polyfunctional molecules.

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